

# MINIMUM INHIBITORY CONCENTRATION OF METRONIDAZOLE AGAINST VACCINAL STRAIN OF *PASTEURELLA MULTOCIDA*

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**ABSTRACT:** Minimum inhibitory concentration was performed against metronidazole that allowed the growth of *Pasteurella multocida* type 1 (B6) but inhibited the growth of *Escherichia coli* which is capable of spoiling vaccine during its production and showed no growth in concentration of 3.2 µg/ml. Metronidazole is an antibiotic of choice in confirming the purity of the cultures of vaccinal strain of *Pasteurella multocida* during vaccine production process.

**Keywords:** *Pasteurella multocida*, Vaccinal strain, MIC, Metronidazole

## INTRODUCTION

*Pasteurella multocida* is Gram negative, nonmotile, facultative anaerobic, coccobacillus. *Pasteurella* infections are usually characterized by local intense cellulitis, purulent discharge, and lymphangitis 12 to 24 hours after the infection [1-3]. The most avoidable mean by which antibiotics become resistant is the misuse of prescribed medication. The proper use of antibiotics should be enforced. Also new products need to be developed including vaccines and diagnostic devices. Most important is that people need to be educated that all the products must be labeled properly for consumers [4]. Minimum inhibitory concentrations against *Pasteurella multocida* isolated from animals was performed. Oxytetracycline, doxycycline, tilmicosin and thiamphenicol for porcine were higher than bovine isolates. MIC of Enrofloxacin was 90 % of the isolates (MIC<sub>90</sub>) of 0.05 µg/ml for isolates from bovine and porcine. Whereas, MIC<sub>90</sub> of > 100 µg/ml was recorded for aminoglycosides [5]. Minimum inhibitory concentrations (MICs) of antibacterial agents against *Mannheimia haemolytica* and *Pasteurella multocida* was investigated and no resistance was detected for ceftiofur and florfenicol. Three strains showed an intermediate susceptibility to tilmicosin. Resistance of *Mannheimia haemolytica* and *Pasteurella multocida* to neomycin, gentamicin, spectinomycin, flumequine, enrofloxacin, and chloramphenicol was from 2% to 16% [6].

Antibiotics can be very well used as identification marker in vaccine production process [7]. The technology can be well exploited to monitor vaccinal strain purity throughout the vaccine production processes. Thus the undesirable microbial species can be eliminated or separated from vaccinal strains. The proposed study envisages antibiogram of *Pasteurella multocida* under varying conditions of growth along with determination of minimal inhibitory concentration (MICs) of selected antibiotics. Our main aim was to determine antibiotics for the purity of culture during vaccine production process.

## MATERIALS AND METHODS

To conduct the study different preliminary steps were taken like sterilization of glassware, media preparation and stains preparation. For this purpose, Nutrient agar, MacConkey's agar, Brain Heart Infusion Agar Muller Hinton Agar, blood

agar and peptone agar were prepared. Vaccinal strain of *Pasteurella multocida* type 1 B6 was obtained from Veterinary Research Institute (VRI) Lahore. The bacteria was inoculated over these media and incubated for 24 hours. After getting the colonies of bacteria, the staining characteristics were studied by Gram's staining; further confirmation of bacterium was conducted by the biochemical tests [8].

### Macrodilution broth susceptibility test

Muller Hinton broth was used for this purpose, medium was prepared in ten flasks, each flask was marked as 0.4 µg/ml, 0.8 µg/ml, 1.6 µg/ml, 3.12 µg/ml, 6.25 µg/ml, 12.5 µg/ml, 25 µg/ml, 50 µg/ml, 100 µg/ml and control (C). The medium was then incubated over night to check sterility. Metronidazole was added in above mentioned nine flasks according to the dilution marked on them and tenth was kept as control. Twenty test tubes measuring 13x100mm, plugged with cotton were placed in hot air oven for 2 hours at 170 °C. These test tubes were grouped into three, each containing ten test tubes. The test tubes were then filled with medium containing antibiotic at the rate of 5ml per tube. Next day single colony from *Pasteurella multocida* and *Escherichia coli* was inoculated into the test tubes of each group. The test tubes were then incubated over night and growth was observed.

## RESULTS AND DISCUSSION

*Pasteurella multocida* was identified by morphological and biochemical characteristic during present study. Minimum Inhibitory Concentration of Metronidazole against *Pasteurella multocida* was performed by serially diluting antibiotic metronidazole as 0.4 µg, 0.8 µg, 1.6 µg/ml, 3.2 µg/ml, 6.4 µg/ml, 12.5 µg/ml, 25 µg/ml, 50 µg/ml and 100 µg/ml. *Pasteurella multocida* type 1(B6) and isolated extraneous contaminant *Escherichia coli* that was identified in this study was inoculated in said dilutions of antibiotic concentrations. *Pasteurella multocida* type 1(B6) grew in all concentrations while contaminant *Escherichia coli* in concentrations of 0.4 µg/ml, 0.8 µg/ml and 1.6 µg/ml, but same *Escherichia coli* showed no growth in concentrations of 3.2 µg/ml, 6.4 µg/ml, 12.5 µg/ml, 25 µg/ml, 50 µg/ml and 100 µg/ml. (Table 1). Minimal inhibitory concentrations of *Pasteurella multocida* were investigated and found highly resistant for spiramycin

and fosfomycin [9]. Whereas, minimum inhibitory concentrations of doxycycline and oxytetracycline against *Pasteurella multocida* recorded MIC of doxycycline 1 µg/ml against *Pasteurella multocida* [10]. Studies on efficacy of antimicrobial agents against *Pasteurella multocida* was investigated by agar diffusion method that showed high sensitivity to various antibiotics including enrofloxacin, chloramphenicol, ampicillin, penicillin G and cephalothin [11]. MICs of spectinomycin and other antimicrobial drugs against *Pasteurella multocida* and *Mannheimia haemolytica* strains from cattle with respiratory infections were performed. Results showed that spectinomycin was highly sensitive strains of *Pasteurella multocida* (93.5%) and *Mannheimia haemolytica* (98.6%) [12]. MIC determination against cefovecin and other reference antimicrobial agents was investigated. The MIC<sub>90</sub> of cefovecin against

*Staphylococcus intermedius* was 0.25 microg/ml, for *Escherichia coli* was 1.0 microg/ml and 0.06 microg/ml for *Pasteurella multocida* [13]. Present study would help in better understanding of the identification procedures for the organisms of *Pasteurella* species, especially *Pasteurella multocida* during laboratory identification and vaccine production.

## CONCLUSIONS

It is concluded that contamination usually occurs during vaccine manufacturing process. The main contaminant was *Escherichia coli* in the present study. *Escherichia coli* were highly susceptible to metronidazole that can be used for purity of the Haemorrhagic septicaemia vaccine

**Table 1. MIC of Metronidazole against Vaccinal strain of *Pasteurella multocida* type 1(B6) and *Escherichia coli*.**

S.No	Species	Concentrations of Metronidazole ( µg per ml of media)									
		0.4	0.	1.6	3.2	6.4	12.5	25	50	100	Control
1.	<i>Pasteurella multocida</i> Type 1(B6)	+	+	+	+	+	+	+	+	+	+
2.	<i>Escherichia coli</i>	+	+	+	-	-	-	-	-	-	+

+ = Growth      - = No growth

## REFERENCES

- [1] Mitnovetski S, Kimble F (2004). Cat bites of the hand. Australian and New Zealand Journal of Surgery. **74**(10):859-62.
- [2] Kwo S, Agarwal JP, Meletiou S (2011). Current treatment of cat bites to the hand and wrist. Journal of Hand Surgery **36**(1):152-3.
- [3] Talan DA, Citron DM, Abrahamian M, Moran GJ, Goldstein EJ. Bacteriologic analysis of infected dog and cat bites (1999). Emergency Medicine Animal Bite Infection Study Group. New England Journal of Medicine. **340**(2):85-92.
- [4] Lewis R (2002). The rise of antibiotic resistant infections. Journal of the American Medical Association **29**(7): 22-30.
- [5] Yoshimura H, Ishimaru M, Endoh YS and Kojima A (2001). Antimicrobial susceptibility of *Pasteurella multocida* isolated from cattle and pigs. Journal of Veterinary Medicine, Series B.48: 555.
- [6] Mevius DJ and Hartman EG (2000). In vitro activity of 12 antibiotics used in veterinary medicine against *Mannheimia haemolytica* and *Pasteurella multocida* isolated from calves in the Netherlands. Tijdschr Diergeneesk. **125**(5):147-152
- [7] Kalhoro D. H, Kalhoro A. B, Rind R, Kalhoro S and Baloch H (2010). In vitro antimicrobial susceptibility of *Pasteurella multocida*. Pakistan Journal of Agriculture, Agricultural Engineering and Veterinary Sciences **26**(2) : 80-86.
- [8] Kalhoro DH, Rind R, Kalhoro MS, Zeb A, Zaman A, Kumbhar S, Mangi MH, Birohi R (2015). Characterization and biochemical behavior assessment of vaccinal strain of *Pasteurella multocida* type 1 b6. International Journal for Agro Veterinary and Medical Sciences, **9**(1):29-35.
- [9] Gutierrez MCB and Ferri EFR (1993). In vitro susceptibility of *Pasteurella multocida* subspecies *multocida* strains isolated from swine to 42 antimicrobial agents. Zentralbl Bakteriologie. **279**(3): 387-393.
- [10] Bousquet E, Morvan H, Aitken I and Hmorgan J (1997). Comparative in vitro activity of doxycycline and oxytetracycline against porcine respiratory pathogens. Veterinary Record. **141** (2): 37-40.
- [11] Harmanssdorfer S and Bauer J (1998). Resistance of bovine and porcine *Pasteurella* to Florfenicol and other antibiotics. Berl Munch Tierarztl. Wochenschr. **111** (11-12):422-426.
- [12] Stefan S, Corinna K, Salmon AS and Watts LJ (2004). In vitro activities of spectinomycin and comparator agents against *Pasteurella multocida* and *Mannheimia haemolytica* from respiratory tract infections of cattle. Journal of Antimicrobial Agents and Chemotherapy. **53**: 379 - 382.
- [13] Stegemann, MR, Passmore CA, Sherington J, Lindeman CJ, Weigel GPDJ and Skoerboe TL (2006). Antimicrobial activity and spectrum of cefovecin, a new extended-spectrum cephalosporin, against pathogens collected from dogs and cats in Europe and North America. Journal of Antimicrobial Agents and Chemotherapy, **50**(7):2286-229